

Nanobiotechnology in the Management of Glaucoma*

Pho Nguyen¹, Alex Huang¹, Samuel C. Yiu^{2,3#}

¹Department of Ophthalmology, Keck School of Medicine of the University of Southern California, Los Angeles, USA; ²The Wilmer Eye Institute, Baltimore, USA; ³King Khaled Eye Specialist Hospital, Riyadh, KSA.
Email: #suiu2@jhmi.edu

Received September 2nd, 2013; revised October 5th, 2013; accepted October 15th, 2013

Copyright © 2013 Pho Nguyen *et al.* This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

ABSTRACT

As the prevalence of glaucoma continues to rise, clinicians and researchers are confronted with an age-old problem: how to reduce risk factors and preserve vision in glaucoma. Current management options revolve around a validated paradigm—*intraocular pressure reduction*. Active investigations to improve drug delivery efficacy and surgical outcomes are flourishing. This article aims to provide the interested readers with a review of recent discoveries in nanobiotechnology for the management of glaucoma. Targeted drug-delivery systems using mesoscale vectors demonstrate promising delivery profiles. The utility of nanoparticulate therapies to support retinal ganglion cell survival is being investigated. Studies to modulate tissue regeneration and remodeling and improve post-trabeculectomy outcomes are underway. Though these modalities promise new avenues to manage glaucoma, immediate market availability is not anticipated soon.

Keywords: Glaucoma; Nanotechnology; Nanomedicine; Drug Delivery; Wound Healing; Nanoparticle

1. Introduction

Glaucoma is a group of diseases having characteristic optic neuropathy with associated visual function deficits. Early disease detection and vision preservation constitute the management of glaucoma. Currently, therapeutic strategies involve medical and surgical reduction of *intraocular pressure (IOP)*. However, maintaining local therapeutic bioavailability of hypotensive agents remains a challenge in the field of ophthalmic drug delivery. Surgical modalities are effective but have associated complications [1]. Given that 60 million people worldwide are afflicted by this disease [2] and 27% of those afflicted are at risk of developing glaucoma-related blindness in one eye after 20 years [3], the demand for viable treatment alternatives, particularly improved drug delivery models, is self-evident.

*Financial Support(s): Alex Huang is supported by the Heed Ophthalmic Foundation.

Pho Nguyen is supported by the Heed Ophthalmic Foundation and the Fletcher Jones Foundation.

Financial Disclosure(s): The authors have no financial interests in the topic of this manuscript. No conflicting relationship exists for any author.

#Corresponding author.

2. Nanobiotechnologies for Glaucoma Management

Previously, many clinical trials demonstrated that initial medical management is an effective option for *IOP reduction* and vision preservation [4-11]. Although topical instillation is the predominant route, ensuring effective local concentration is a fundamental challenge. Precorneal factors such as small load, poor tissue penetration, mechanical removal by tears and blinking, and nasolacrimal elimination prevent the medication from reaching a sustained therapeutic level and necessitate multiple dosings per day, ultimately, leading to poor patient adherence [12-14]. Other studies further report that many patients waste much of the topical medications because of incorrect instillation techniques [13-15]. Systemic absorption may precipitate adverse effects [16-21]. Therefore, efforts are directed toward developing better drug delivery systems, e.g. ophthalmic inserts, smart hydrogel, vesicular and particulate carrier systems.

Nanoscale drug delivery systems offer the therapeutic advantages of targeted tissue penetration, enhanced release kinetics, and increased local biodistribution [22-24]. In the field of tissue engineering and regenerative medi-

cine, the biologic length scale of nanostructures promotes meaningful interactions for tissue repair and regeneration [23]. “Nano” was originally defined for the semiconductor industry as having at least one dimension less than 100 nanometer; but the definition of “nano-” has been recast in an operational fashion and many authors use “nano-” to include biologically relevant length scale, such as larger macromolecules and organelles for the fields of nanobiotechnology and nanomedicine [22-24]. Below, we summarize the recent discoveries in nanobiotechnology for the management of glaucoma.

2.1. Drug Delivery Systems

2.1.1. Vesicular Platforms—Liposomes, Discomes, and Niosomes

Modeled after biologic organelles, liposomal systems are designed for encapsulation of therapeutic agents for drug delivery and have found to have increased efficacy and decreased toxicity. Their diverse systemic applications encompasses liposomal daunorubicin for Kaposi’s sarcoma, micellar paclitaxel for ovarian and breast cancer, and liposomal amphotericin B for systemic fungal infections or leishmaniasis. Surface modifications, such as PEGylation of the liposome, extend systemic circulating time and reduce toxicity.

Liposomal preparations of topical ophthalmic medications are particularly well studied for ocular surface infections. Use of liposomal amphotericin B for the treatment of fungal keratitis has been reported to significantly reduce toxicity compared to non-liposomal formulation [25,26]. Topical liposomal acyclovir shows better delivery than commercial acyclovir ointment in *in vitro* studies and animal models [27]. Liposomal formulations of gatifloxacin, ciprofloxacin and fluconazole have shown prolonged delivery profiles, leading to better *in vitro* transcorneal permeation compared to aqueous formulations [28-30].

Other investigators are evaluating the feasibility of liposome-encapsulated ocular antihypertensive drugs. Topical liposomal and niosomal formulations of acetazolamide, a carbonic anhydrase inhibitor, are being investigated for their ability to overcome acetazolamide’s limited aqueous solubility and improve its corneal permeation [31-34]. *In vitro* and *in vivo* studies demonstrate sustained release kinetics with good intraocular hypotensive effect [32].

In addition to liposomes, other vesicular platforms, such as discomes and niosomes, are also being explored for ocular antihypertensive medications. Discomes are non-ionic surfactant-based discoidal vesicles, which also improve drug delivery. Some authors have reported that discoidal vesicles containing timolol maleate produce a sustained activity profile upon introduction to the ocular

cavity [35]. Niosomes are another non-ionic bilayered vesicle that can entrap both hydrophilic and lipophilic drugs, either in the aqueous layer or in the vesicular membrane. Compared to liposomes, niosomes offer some advantages, such as chemical stability and lower production cost. A recent study reported that a mucoadhesive-coated niosomal system for timolol maleate can achieve higher peak concentrations for an extended period compared to the conventional dose form [36]. Sizes of these discomes and niosomes are in the micron range [35,36]. Lastly, a more recent nanovesicular formulation of brimonidine tartrate has been constructed using sorbitan stearate and cholesterol, and the authors report significant intraocular pressure-lowering activity for a prolonged period of time compared to the commercial preparation [37-39]. Most of the above studies report no short-term toxicity to the ocular surface of the animal models.

2.1.2. Nanoparticulate Platforms

Development of nanoparticulate platforms for drug delivery has gained significant traction over recent years. The nanoscale size range and surface functionalization offer the advantages of targeted delivery and resistance to degradation. Particulate delivery systems also provide more stable storage media, compared to vesicular systems. Biodegradable polymeric platforms present an exciting opportunity for investigators, where the pharmacokinetics is determined by the particulate size and carrier material, which is ultimately controlled by synthesis technologies.

Biodegradable poly(lactic-co-glycolic) acid (PLGA) polymers have promise as potential couriers for antihypertensive agents. Timolol maleate integrated into PLGA polymers via a solvent evaporation method has been reported to have sustained release kinetics [40]. The release mechanism is proposed to be polymeric degradation, rather than microporous diffusion. Another study suggests that PLGA and poly(l-lactide) acid (PLLA) micro- and nanoparticles can deliver timolol maleate continually over a 3-month period [41]. Intravitreal injection of PLGA is safe in a rabbit model, where only a localized foreign body reaction is reported [42]. In this study, the authors observed that the choroid and retina maintain normal appearance and no clinical inflammation was detected. Accordingly, some investigators are optimistic that these biodegradable polymeric micro- and nanospheres may find application as subconjunctival depots to improve patient adherence [41]. Similarly, nanoparticulate platforms for carbonic anhydrase inhibitors are being investigated as well. Methazolamide nanoparticles are reported to have longer and higher therapeutic efficacy compared to suspension formulation and commercial eye drops [43,44].

Another polymer of ophthalmic interest is chitosan, a polycationic biodegradable polymer composed of repeating glucosamine units. The chitosan nanoparticles are thought to have longer precorneal residence time and increased corneal penetration. Gatifloxacin-loaded chitosan nanoparticles show sustained released kinetics [45]. Chitosan/poly(lactic acid) nanoparticles containing rapamycin demonstrate improved corneal allograft survival in a rabbit model compared to aqueous suspension [46]. With respect to ocular anti-hypertensive agents, nanoparticulate formulations of chitosan and timolol maleate or dorzolamide hydrochloride have also been described [47]. These investigators find that modification of these nanoparticles with hyaluronic acid, which improves mucoadhesion, significantly decreases intraocular pressure in a rabbit model, compared to plain solutions of these drugs.

2.1.3. Dendrimeric Platforms

Dendrimers comprise a fascinating drug carrier system. These hyperbranched globular macromolecules with dendritic subunits have modifiable properties depending on their subunits and surface terminal groups. Therapeutic agents can either be encapsulated in the core of the dendrimer scaffold or conjugated on the surface. The dendrimer diameter can be customized based on its generation and terminal groups.

Some therapeutic agents being investigated include propranolol, sulfasalazine, folic acid, adriamycin, methotrexate, paclitaxel, and penicillin. Photodynamic therapy and gene therapy using dendrimers for corneal, retinal, and choroidal neovascularization have been published [48-50]. For glaucoma, poly(amidoamine) dendrimeric constructs have been constructed to deliver pilocarpine and tropicamide [51]. These dendrimers appear to have greater corneal residence time, increased bioavailability, and low ocular irritation index in animal model. A recent study reported increased uptake of timolol maleate and brimonidine using poly(amidoamine) dendrimer hydrogel as delivery modality in bovine corneal model [52].

2.2. Tissue Protection

Currently, IOP control drives glaucoma management. However, some investigators are evaluating the feasibility of neuroprotection as well. Apoptosis of retinal ganglion cells (RGC) has been associated with glaucomatous optic neuropathy [53]. Studies using RGC death after optic nerve injury in an animal model suggest that neurotrophic factors, e.g. brain-derived growth factors, insulin-like growth factors, glial-derived neurotrophic factor, and ciliary neurotrophic factors (CNTF) may support RGC survival [54-56]. Accordingly, the roles of neurotrophic factors in neuroprotection in glaucoma are being

explored. Lentiviral-mediated transfer of CNTF into Schwann cells for optic nerve repair has been found to significantly increase RGC survival in animal models [57]. Others investigators have used biodegradable PLGA polymers to construct nano- and microspheres for CNTF encapsulation [58]. These authors report bioactivity in an *in vitro* neural stem cell model and note that the process of protein encapsulation with the polymer did not reduce its potency. Others incorporate CNTF nanospheres into photopolymerizable hydrogel to create a tissue engineering scaffold with intrinsic sustained drug delivery capability [59]. This scaffold provides an enhanced substratum for neural tissue repair and regeneration. Recently, induction of heat shock protein for neuroprotection using superparamagnetic nanoparticles has also been reported [60]. In the near future, successful commercial deployment of these and other developments will add another set of tools in the armamentarium of the glaucoma specialist to preserve vision.

2.3. Modulation of Postsurgical Wound Healing in Filtration Surgery

Successful glaucoma filtering surgery necessitates adequate passage of the aqueous humor from the anterior chamber to an extraocular reservoir. Exuberant cicatricial changes to the conjunctiva following glaucoma filtration procedures present a threat to long-term success [61,62]. Antifibrotic agents have been demonstrated to promote longer bleb survival but are associated with severe complications such as leakage, infection, hypotony, and endophthalmitis [63-65]. Currently, antifibrotic agents are mostly frequently introduced intraoperatively via a soaked sponge. As with drug delivery of intraocular pressure lowering medications, nanobiotechnology can be directed toward improvement of bleb survival.

A disc carrying biodegradable poly(lactic acid) microspheres loaded with 5-fluorouracil for subconjunctival implant has been developed and tested in a rabbit model [66]. The authors show that delivery of 5-fluorouracil using the carrier resulted in greater decrease in intraocular pressure, prolonged bleb persistence, and less corneal toxicity. Syntheses of various nanoparticulate 5-fluorouracil and mitomycin have also been reported [67-69]. However, ophthalmic applications of these nanoparticles have yet to be investigated.

Other investigators target the growth factor mediated cellular proliferation. In one study, glucosamine and glucosamine 6-sulfate dendrimers were used to target fibroblast growth factor-2 mediated endothelial cell proliferation and neoangiogenesis in a rabbit model of scar tissue formation after glaucoma filtration surgery [70]. The authors report that postoperative day 30 bleb survival improves from 30% to 80%. Histologic examination revealed less cicatricial proliferation in the den-

dimer group. In another approach, biodegradable porous PLGA microspheres containing antisense-TGF- β 2 oligonucleotide nanocomplexes have been found to promote bleb survival in a rabbit model of filtering surgery [71]. The antisense oligonucleotide reduces synthesis of the cytokine TGF- β 2, which promotes wound healing. In this study, the nanocomplexes are encapsulated into porous microspheres and administered subconjunctivally. The authors report increased penetration of the encapsulated oligonucleotides in conjunctival cells and increased time to filtering bleb failure. Steroids may also be packaged via nanoparticles to modulate postoperative wound healing. Dexamethasone entrapped in biodegradable poly(d,l-lactide-co-glycolide) (PLGA) nanoparticles has been reported [72]. These nanoparticles can find applications both in preventing filtering bleb scarring and in the investigation of steroid response in glaucoma.

Viral vectors constitute another form of particulate nanoparticles that can be directed toward glaucoma surgical management. Adenovirus-mediated gene therapy has been performed to prevent bleb scarring in a rabbit model of glaucoma filtration surgery [73]. The authors find that topical, intraoperative application of recombinant adenovirus containing the human p21 gene demonstrates inhibition of wound healing and fibroproliferation after filtration surgery, comparable to mitomycin but with fewer adverse effects. Similar results are seen with an Ad-p27 vector [74]. Interestingly, attempts to use viral vectors to blunt steroid response are also underway. Novel glucocorticoid-inducible adenovirus vectors have been developed to overproduce metalloprotein 1, which degrades collagen type I after specific activation by dexamethasone [75]. Therefore, patients who presumably need long-term steroid treatment may benefit by having increased metalloproteinase activity to aid trabecular flow while on steroids.

3. Conclusion

This brief review highlights the recent advances and advantages of nanobiotechnology for improving our understanding of glaucoma and its management. In essence, nanobiotechnology offers the potential for more effective delivery of pharmaceutical agents that can influence both the medical and surgical arms of glaucoma management.

REFERENCES

- [1] S. J. Gedde, J. C. Schiffman, W. J. Feuer, *et al.*, (Tube versus Trabeculectomy Study Group), "Three-Year Follow-Up of the Tube versus Trabeculectomy Study," *American Journal of Ophthalmology*, Vol. 148, No. 5, 2009, pp. 670-684. <http://dx.doi.org/10.1016/j.ajo.2009.06.018>
- [2] H. A. Quigley, "Glaucoma," *Lancet*, Vol. 377, No. 9774, 2011, pp. 1367-1377. [http://dx.doi.org/10.1016/S0140-6736\(10\)61423-7](http://dx.doi.org/10.1016/S0140-6736(10)61423-7)
- [3] M. G. Hattenhauer, D. H. Johnson, H. H. Ing, *et al.*, "The Probability of Blindness from Open-Angle Glaucoma," *Ophthalmology*, Vol. 105, No. 9, 1998, pp. 2099-2104. [http://dx.doi.org/10.1016/S0161-6420\(98\)91133-2](http://dx.doi.org/10.1016/S0161-6420(98)91133-2)
- [4] M. A. Kass, D. K. Heuer, E. J. Higginbotham, *et al.*, (For the Ocular Hypertension Treatment Study Group), "A Randomized Trial Determines That Topical Ocular Hypotensive Medication Delays or Prevents the Onset of Primary Open-Angle Glaucoma," *Archives of Ophthalmology*, Vol. 120, No. 6, 2002, pp. 701-713. <http://dx.doi.org/10.1001/archophth.120.6.701>
- [5] S. Miglior, T. Zeyen, N. Pfeiffer, *et al.*, "Results of the European Glaucoma Prevention Study," *Ophthalmology*, Vol. 112, No. 9, 2005, pp. 366-375. <http://dx.doi.org/10.1016/j.ophtha.2005.06.020>
- [6] J. Burr, A. Azuara-Blanco and A. Avenell, "Medical versus Surgical Interventions for Open Angle Glaucoma," *Cochrane Database of Systematic Reviews*, Vol. 18, 2005, Article ID: CD004399.
- [7] M. C. Leske, A. Heijl, L. Hyman, *et al.*, "Predictors of Long-Term Progression in the Early Manifest Glaucoma Trial," *Ophthalmology*, Vol. 114, No. 11, 2007, pp. 1965-1972. <http://dx.doi.org/10.1016/j.ophtha.2007.03.016>
- [8] M. A. Kass, M. O. Gordon, F. Gao, *et al.*, (For the Ocular Hypertension Treatment Study Group), "Delaying Treatment of Ocular Hypertension," *Archives of Ophthalmology*, Vol. 128, No. 3, 2010, pp. 276-287. <http://dx.doi.org/10.1001/archophth.2010.20>
- [9] A. C. S. Crichton, P. Harasymowycz, C. M. L. Hutnik, *et al.*, "Effectiveness of Dorzolamide-Timolol (Cosopt) in Patients Who Were Treatment Naive for Open-Angle Glaucoma or Ocular Hypertension: The COSOPT First-Line Study," *Journal of Ocular Pharmacology and Therapeutics*, Vol. 26, No. 5, 2010, pp. 503-511. <http://dx.doi.org/10.1089/jop.2010.0032>
- [10] D. C. Musch, B. W. Gillespie, P. R. Lichter, *et al.*, "Visual Field Progression in the Collaborative Initial Glaucoma Treatment Study: The Impact of Treatment and Other Baseline Factors," *Ophthalmology*, Vol. 116, No. 2, 2009, pp. 200-207. <http://dx.doi.org/10.1016/j.ophtha.2008.08.051>
- [11] L. J. Katz, W. C. Steinmann, A. Kabir, *et al.*, "Selective Laser Trabeculectomy versus Medical Therapy as Initial Treatment of Glaucoma: A Prospective, Randomized Trial," *Journal of Glaucoma*, Vol. 21, No. 7, 2011, pp. 460-468.
- [12] N. M. Davies, "Biopharmaceutical Considerations in Topical Ocular Drug Delivery," *Clinical and Experimental Pharmacology and Physiology*, Vol. 27, No. 7, 2000, pp. 558-562. <http://dx.doi.org/10.1046/j.1440-1681.2000.03288.x>
- [13] T. Tsai, A. L. Robin and J. P. Smith III, "An Evaluation of How Glaucoma Patients Use Topical Medications: A Pilot Study," *Transactions of the American Ophthalmological Society*, Vol. 105, 2007, pp. 29-33.
- [14] C. M. Olthoff, J. G. Hoevenaars, B. W. van den Borne, *et al.*, "Prevalence and Determinants of Non-Adherence to Topical Hypotensive Treatment in Dutch Glaucoma Patients," *Graefe's Archive for Clinical and Experimental*

- Ophthalmology*, Vol. 247, No. 2, 2009, pp. 235-243.
<http://dx.doi.org/10.1007/s00417-008-0944-y>
- [15] R. Gupta, B. Patil, B. M. Shah, *et al.*, "Evaluating Eye Drop Instillation Technique in Glaucoma Patients," *Journal of Glaucoma*, Vol. 21, No. 3, 2012, pp. 189-192.
<http://dx.doi.org/10.1097/IJG.0b013e31820bd2e1>
- [16] F. C. Hugues and C. Le Jeunne, "Systemic and Local Tolerability of Ophthalmic Drug Formulations. An Update," *Drug Safety*, Vol. 8, No. 5, 1993, pp. 365-380.
<http://dx.doi.org/10.2165/00002018-199308050-00004>
- [17] G. Walters and R. H. Taylor, "Severe Systemic Toxicity Caused by Brimonidine Drops in an Infant with Presumed Juvenile Xanthogranuloma," *Eye*, Vol. 13, 1999, pp. 797-798.
<http://dx.doi.org/10.1038/eye.1999.235>
- [18] J. O. Carlsen, N. A. Zabriskie, Y. H. Kwon, *et al.*, "Apparent Central Nervous System Depression in Infants after the Use of Topical Brimonidine," *American Journal of Ophthalmology*, Vol. 128, No. 2, 1999, pp. 255-256.
[http://dx.doi.org/10.1016/S0002-9394\(99\)00083-5](http://dx.doi.org/10.1016/S0002-9394(99)00083-5)
- [19] R. J. Bowman, J. Cope and K. K. Nischal, "Ocular and Systemic Side Effects of Brimonidine 0.2% Eye Drops (Alphagan) in Children," *Eye*, Vol. 18, 2004, pp. 24-26.
<http://dx.doi.org/10.1038/sj.eye.6700520>
- [20] A. P. Demayo and M. M. Reidenberg, "Grand Mal Seizure in a Child 30 Minutes after Cyclogyl (Cyclopentolate Hydrochloride) and 10% Neo-Synephrine (Phenylephrine Hydrochloride) Eye Drops Were Instilled," *Pediatrics*, Vol. 113, No. 5, 2004, pp. e499-e500.
<http://dx.doi.org/10.1542/peds.113.5.e499>
- [21] R. F. Wang, D. J. Gagliuso and S. M. Podos, "Effect of Flunarizine, a Calcium Channel Blocker, on Intraocular Pressure and Aqueous Humor Dynamics in Monkeys," *Journal of Glaucoma*, Vol. 17, No. 1, 2008, pp. 73-78.
<http://dx.doi.org/10.1097/IJG.0b013e318133a845>
- [22] O. M. Koo, I. Rubinstein and H. Onyuksel, "Role of Nanotechnology in Targeted Drug Delivery and Imaging: A Concise Review," *Nanomedicine*, Vol. 1, No. 3, 2005, pp. 193-212.
<http://dx.doi.org/10.1016/j.nano.2005.06.004>
- [23] P. Nguyen, M. Meyyappan and S. C. Yiu, "Applications of Nanobiotechnology in Ophthalmology—Part I," *Ophthalmic Research*, Vol. 44, 2010, pp. 1-16.
<http://dx.doi.org/10.1159/000279436>
- [24] R. A. Petros and J. M. DeSimone, "Strategies in the Design of Nanoparticles for Therapeutic Applications," *Nature Reviews Drug Discovery*, Vol. 9, 2010, pp. 615-627.
<http://dx.doi.org/10.1038/nrd2591>
- [25] J. P. Adler-Moore and R. T. Proffitt, "Development, Characterization, Efficacy and Mode of Action of Ambisome, a Unilamellar Liposome Formulation of Amphotericin B," *Journal of Liposome Research*, Vol. 3, No. 3, 1993, pp. 429-450.
<http://dx.doi.org/10.3109/08982109309150729>
- [26] K. Morand, A. C. Bartoletti, A. Bochet, *et al.*, "Liposomal Amphotericin B Eye Drops to Treat Fungal Keratitis: Physico-Chemical and Formulation Stability," *International Journal of Pharmaceutics*, Vol. 344, No. 1-2, 2007, pp. 150-153.
<http://dx.doi.org/10.1016/j.ijpharm.2007.04.028>
- [27] P. Chetoni, S. Rossi, S. Buralassi, *et al.*, "Comparison of Liposome-Encapsulated Acyclovir with Acyclovir Ointment: Ocular Pharmacokinetics in Rabbits," *Journal of Ocular Pharmacology and Therapeutics*, Vol. 20, No. 2, 2004, pp. 169-177.
<http://dx.doi.org/10.1089/108076804773710849>
- [28] K. M. Hosny, "Optimization of Gatifloxacin Liposomal Hydrogel for Enhanced Transcorneal Permeation," *Journal of Liposome Research*, Vol. 20, No. 1, 2010, pp. 31-37.
<http://dx.doi.org/10.3109/08982100903030255>
- [29] K. M. Hosny, "Ciprofloxacin as Ocular Liposomal Hydrogel," *AAPS PharmSciTech*, Vol. 11, No. 1, 2010, pp. 241-246.
<http://dx.doi.org/10.1208/s12249-009-9373-4>
- [30] F. S. Habib, E. A. Fouad, M. S. Abdel-Rhman, *et al.*, "Liposomes as an Ocular Delivery System of Fluconazole: In-Vitro Studies," *Acta Ophthalmologica*, Vol. 88, No. 8, 2010, pp. 901-904.
<http://dx.doi.org/10.1111/j.1755-3768.2009.01584.x>
- [31] O. N. El-Gazayerly and A. H. Hikal, "Preparation and Evaluation of Acetazolamide Liposomes as an Ocular Delivery System," *International Journal of Pharmaceutics*, Vol. 158, No. 2, 1997, pp. 121-127.
[http://dx.doi.org/10.1016/S0378-5173\(97\)00186-5](http://dx.doi.org/10.1016/S0378-5173(97)00186-5)
- [32] D. Aggarwal, A. Garg and I. P. Kaur, "Development of a Topical Niosomal Preparation of Acetazolamide: Preparation and Evaluation," *Journal of Pharmacy and Pharmacology*, Vol. 56, No. 12, 2004, pp. 1509-1517.
<http://dx.doi.org/10.1211/0022357044896>
- [33] A. S. Guinedi, N. D. Mortada, S. Mansour, *et al.*, "Preparation and Evaluation of Reverse-Phase Evaporation and Multilamellar Niosomes as Ophthalmic Carriers of Acetazolamide," *International Journal of Pharmaceutics*, Vol. 306, No. 1-2, 2005, pp. 71-82.
<http://dx.doi.org/10.1016/j.ijpharm.2005.09.023>
- [34] R. M. Hathout, S. Mansour, N. D. Mortada, *et al.*, "Liposomes as an Ocular Delivery System for Acetazolamide: in Vitro and in Vivo Studies," *AAPS PharmSciTech*, Vol. 8, No. 1, 2007, pp. E1-E12.
<http://dx.doi.org/10.1208/pt0801001>
- [35] S. P. Vyas, N. Mysore, V. Jaitely, *et al.*, "Discoidal Niosome Based Controlled Ocular Delivery of Timolol Maleate," *Pharmazie*, Vol. 53, No. 7, 1998, pp. 466-469.
- [36] I. P. Kaur, D. Aggarwal, H. Singh and S. Kakkar, "Improved Ocular Absorption Kinetics of Timolol Maleate Loaded into a Bioadhesive Niosomal Delivery System," *Graefe's Archive for Clinical and Experimental Ophthalmology*, Vol. 248, No. 10, 2010, pp. 1467-1472.
<http://dx.doi.org/10.1007/s00417-010-1383-0>
- [37] P. Prabhu, K. R. Nitish, M. Koland, N. Harish, K. Vijayanarayan, G. Dhondge and R. Charyulu, "Preparation and Evaluation of Nano-Vesicles of Brimonidine Tartrate as an Ocular Drug Delivery System," *Journal of Young Pharmacists*, Vol. 2, No. 4, 2010, pp. 356-361.
<http://dx.doi.org/10.4103/0975-1483.71623>
- [38] S. Maiti, S. Paul, R. Mondol, S. Ray and B. Sa, "Nanovesicular formulation of Brimonidine Tartrate for the Management of Glaucoma: In Vitro and in Vivo Evaluation," *AAPS PharmSciTech*, Vol. 12, No. 2, 2011, pp. 755-763.
<http://dx.doi.org/10.1208/s12249-011-9643-9>

- [39] M. Sabyasachi, P. Sayon, M. Ranjit, R. Somasree and S. Biawanath, "Nanovesicular Formulation of Brimonidine Tartrate for the Management of Glaucoma: *In Vitro* and *in Vivo* Evaluation," *AAPS PharmSciTech*, Vol. 12, No. 2, 2011, pp. 755-763.
<http://dx.doi.org/10.1208/s12249-011-9643-9>
- [40] C. Stureson, J. Carlfors, K. Edsman and M. Andersson, "Preparation of Biodegradable Poly(lactic-co-glycolic) Acid Microspheres and Their *in Vitro* Release of Timolol Maleate," *International Journal of Pharmaceutics*, Vol. 89, No. 3, 1993, pp. 235-244.
[http://dx.doi.org/10.1016/0378-5173\(93\)90249-F](http://dx.doi.org/10.1016/0378-5173(93)90249-F)
- [41] J. P. Bertram, S.S. Saluja, J. McKain and E. B. Lavik, "Sustained Delivery of Timolol Maleate from Poly(lactic-co-glycolic acid)/Poly(lactic acid) Microspheres for over 3 Months," *Journal of Microencapsulation*, Vol. 26, No. 1, 2009, pp. 18-26.
<http://dx.doi.org/10.1080/02652040802095250>
- [42] G. G. Giordano, P. Chevez-Barrios, M. F. Refojo and C. A. Garcia, "Biodegradation and Tissue Reaction to Intravitreal Biodegradable Poly(D,L-lactic-co-glycolic)-Acid Microsphere," *Current Eye Research*, Vol. 14, No. 9, 1995, pp. 761-768.
<http://dx.doi.org/10.3109/02713689508995797>
- [43] R. Chen, Y. Qian, R. Li, Q. Zhang, D. F. Liu, M. Wang and Q. W. Xu, "Methazolamide Calcium Phosphate Nanoparticles in An Ocular Delivery System," *Yakugaku Zasshi*, Vol. 130, No. 3, 2010, pp. 419-424.
<http://dx.doi.org/10.1248/yakushi.130.419>
- [44] R. Li, S. Jiang, D. Liu, X. Y. Bi, F. Z. Wang, Q. Zhang and Q. W. Xu, "A Potential New Therapeutic System for Glaucoma: Solid Lipid Nanoparticles Containing Methazolamide," *Journal of Microencapsulation*, Vol. 28, No. 2, 2011, pp. 134-141.
<http://dx.doi.org/10.3109/02652048.2010.539304>
- [45] S. K. Motwani, S. Chopra, S. Talegaonkar, K. Kohli, F. J. Ahmad and R. K. Khar, "Chitosan-Sodium Alginate Nanoparticles as Submicroscopic Reservoirs for Ocular Delivery: Formulation, Optimisation and *in Vitro* Characterization," *European Journal of Pharmaceutics and Biopharmaceutics*, Vol. 68, No. 3, 2008, pp. 513-525.
- [46] X. B. Yuan, Y. B. Yuan, W. Jiang, *et al.*, "Preparation of Rapamycin-Loaded Chitosan/PLA Nanoparticles for Immunosuppression in Corneal Transplantation," *International Journal of Pharmaceutics*, Vol. 349, No. 1-2, 2008, pp. 241-248.
<http://dx.doi.org/10.1016/j.ijpharm.2007.07.045>
- [47] S. Wadhwa, R. Paliwal, S. R. Paliwal and S. P. Vyas, "Hyaluronic Acid Modified Chitosan Nanoparticles for Effective Management of Glaucoma: Development, Characterization, and Evaluation," *Journal of Drug Targeting*, Vol. 18, No. 4, 2010, pp. 292-302.
<http://dx.doi.org/10.3109/10611860903450023>
- [48] R. J. Marano, N. Wimmer, P. S. Kearns, B. G. Thomas, I. Toth, M. Brankov and P. E. Rakoczy, "Inhibition of *in Vitro* VEGF Expression and Choroidal Neovascularization by Synthetic Dendrimer Peptide Mediated Delivery of a Sense Oligonucleotide," *Experimental Eye Research*, Vol. 79, No. 4, 2004, pp. 525-535.
<http://dx.doi.org/10.1016/j.exer.2004.06.023>
- [49] R. Ideta, F. Tasaka, W. D. Jang, *et al.*, "Nanotechnology-Based Photodynamic Therapy for Neovascular Disease using a Supramolecular Nanocarrier Loaded with a Dendritic Photosensitizer," *Nano Letters*, Vol. 5, No. 12, 2005, pp. 2426-2431.
<http://dx.doi.org/10.1021/nl051679d>
- [50] K. Sugisaki, T. Usui, N. Nishiyama, W. D. Jang, Y. S. Yanagi, S. Yamagami, S. Amano and K. Kataoka, "Photodynamic Therapy for Corneal Neovascularization Using Polymeric Micelles Encapsulating Dendrimer Porphyrins," *Investigative Ophthalmology & Visual Science*, Vol. 49, No. 3, 2008, pp. 894-899.
<http://dx.doi.org/10.1167/iovs.07-0389>
- [51] Th. F. Vandamme and L. Brobeck, "Poly(amidoamine) Dendrimers as Ophthalmic Vehicles for Ocular Delivery of Pilocarpine Nitrate and Tropicamide," *Journal of Controlled Release*, Vol. 102, No. 1, 2005, pp. 23-38.
<http://dx.doi.org/10.1016/j.jconrel.2004.09.015>
- [52] C. A. Holden, P. Tyagi, A. Thakur, R. Kadam, G. Jadhav, U. B. Kompella and H. Yang, "Polyamidoamine Dendrimer Hydrogel for Enhanced Delivery of Antiglaucoma Drugs," *Nanomedicine*, Vol. 8, No. 5, 2012, pp. 776-783.
- [53] H. A. Quigley, R. W. Nickells, L. A. Kerrigan, M. E. Pease, D. J. Thibault and D. J. Zack, "Retinal Ganglion Cell Death in Experimental Glaucoma and after Axotomy Occurs by Apoptosis," *Investigative Ophthalmology & Visual Science*, Vol. 36, No. 5, 1995, pp. 774-786.
- [54] J. Weise, S. Isenmann, N. Klöcker, S. Kugler, S. Hirsch, C. Gravel and M. Bahr, "Adenovirus-Mediated Expression of Ciliary Neurotrophic Factor (CNTF) Rescues Axotomized Rat Retinal Ganglion Cells but Does Not Support Axonal Regeneration *in Vivo*," *Neurobiology of Disease*, Vol. 7, No. 3, 2000, pp. 212-223.
<http://dx.doi.org/10.1006/nbdi.2000.0285>
- [55] G. Parrilla-Reverter, M. Agudo, P. Sobrado-Calvo, *et al.*, "Effects of Different Neurotrophic Factors on the Survival of Retinal Ganglion Cells after a Complete Intraocular Nerve Crush Injury: A Quantitative *in Vivo* Study," *Experimental Eye Research*, Vol. 89, No. 1, 2009, pp. 32-41.
<http://dx.doi.org/10.1016/j.exer.2009.02.015>
- [56] M. D. Pease, D. J. Zack, C. Berlinicke, *et al.*, "Effect of CNTF on Retinal Ganglion Cell Survival in Experimental Glaucoma," *Investigative Ophthalmology & Visual Science*, Vol. 50, No. 5, 2009, pp. 2194-2200.
<http://dx.doi.org/10.1167/iovs.08-3013>
- [57] Y. Hu, S. G. Leaver, G. W. Plant, W. T. J. Hendriks, S. P. Niclou, J. Verhaagen, A. R. Harvey and Q. Cui, "Lentiviral-Mediated Transfer of CNTF to Schwann Cells within Reconstructed Peripheral Nerve Grafts Enhances Adult Retinal Ganglion Cell Survival and Axonal Regeneration," *Molecular Therapy*, Vol. 11, No. 6, 2005, pp. 906-915.
<http://dx.doi.org/10.1016/j.ymthe.2005.01.016>
- [58] M. K. Nkansah, S. Y. Tzeng, A. M. Holdt and E. B. Lavik, "Poly(lactic-co-glycolic acid) Nanospheres and Microspheres for Short and Long-Term Delivery of Bioactive Ciliary Neurotrophic Factor," *Biotechnology and Bioengineering*, Vol. 100, No. 5, 2008, pp. 1010-1019.
<http://dx.doi.org/10.1002/bit.21822>
- [59] S. Y. Tzeng and E. B. Lavik, "Photopolymerizable Nanonarray Hydrogels Deliver CNTF and Promote Differen-

- tiation of Neural Stem Cells,” *Soft Matter*, Vol. 6, No. 10, 2010, pp. 2208-2215. <http://dx.doi.org/10.1039/b923544b>
- [60] M. Jeun, J. W. Jeoung, S. Moon, *et al.*, “Engineered Superparamagnetic $Mn_{0.5}Zn_{0.5}Fe_2O_4$ Nanoparticles as a Heat Shock Protein Induction Agent for Ocular Neuroprotection in Glaucoma,” *Biomaterials*, Vol. 32, No. 2, 2011, pp. 387-394. <http://dx.doi.org/10.1016/j.biomaterials.2010.09.016>
- [61] G. L. Skuta and R. K. Parrish 2nd, “Wound Healing in Glaucoma Filtering Surgery,” *Survey of Ophthalmology*, Vol. 32, No. 3, 1987, pp. 149-170.
- [62] M. M. Tahery and D. A. Lee, “Pharmacologic Control of Wound Healing in Glaucoma Filtration Surgery,” *Journal of Ocular Pharmacology and Therapeutics*, Vol. 5, No. 2, 1989, pp. 155-179. <http://dx.doi.org/10.1089/jop.1989.5.155>
- [63] D. A. Morris, M. O. Peracha, D. H. Shin, C. Kim, S. C. Cha and Y. Y. Kim, “Risk Factors for Early Filtration Failure Requiring Suture Release after Primary Glaucoma Triple Procedure with Adjunctive Mitomycin,” *JAMA Ophthalmology*, Vol. 117, No. 9, 1999, pp. 1149-1154. <http://dx.doi.org/10.1001/archophth.117.9.1149>
- [64] P. W. DeBry, T. W. Perkins, G. Heatley, P. Kaufman, ; L. C. Brumback, “Incidence of Late-Onset Bleb-Related Complications Following Trabeculectomy with Mitomycin,” *JAMA Ophthalmology*, Vol. 120, No. 3, 2002, pp. 297-300. <http://dx.doi.org/10.1001/archophth.120.3.297>
- [65] A. M. Palanca-Capistrano, J. Hall, L. B. Cantor, L. Morgan, J. Hoop and D. Wudunn, “Long-Term Outcomes of Intraoperative 5-Fluorouracil versus Intraoperative Mitomycin C in Primary Trabeculectomy Surgery,” *Ophthalmology*, Vol. 116, No. 2, 2009, pp. 185-190. <http://dx.doi.org/10.1016/j.ophtha.2008.08.009>
- [66] L. J. Cui, N. X. Sun, X. H. Li, J. Huang and J. G. Yang, “Subconjunctival Sustained Release 5-Fluorouracil for Glaucoma Filtration Surgery,” *Acta Pharmacologica Sinica*, Vol. 29, No. 9, 2008, pp. 1021-1028. <http://dx.doi.org/10.1111/j.1745-7254.2008.00833.x>
- [67] Z. Hou, H. Wei, Q. Wang, *et al.*, “New Method to Prepare Mitomycin C Loaded PLA-Nanoparticles with High Drug Entrapment Efficiency,” *Nanoscale Research Letters*, Vol. 4, No. 7, 2009, pp. 732-737. <http://dx.doi.org/10.1007/s11671-009-9312-z>
- [68] A. E. Yassin, M. K. Anwer, H. A. Mowafy, I. M. El-Bagory, M. A. Bayomi and I. A. Alsarra, “Optimization of 5-Fluorouracil Solid-Lipid Nanoparticles: A Preliminary Study to Treat Colon Cancer,” *International Journal of Medical Sciences*, Vol.7, No. 6, 2010, pp. 398-408. <http://dx.doi.org/10.7150/ijms.7.398>
- [69] A. J. Shuhendler, R. Y. Cheung, J. Manias, A. Connor, A. M. Rauth and X. Y. Wu, “A Novel Doxorubicin-Mitomycin C Co-Encapsulated Nano-Particle Formulation Exhibits Anti-Cancer Synergy in Multidrug Resistant Human Breast Cancer Cells,” *Breast Cancer Research and Treatment*, Vol. 119, No. 2, 2010, pp. 255-269. <http://dx.doi.org/10.1007/s10549-008-0271-3>
- [70] S. Shaunak, S. Thomas, E. Gianasi, *et al.*, “Polyvalent Dendrimer Glucosamine Conjugates Prevent Scar Tissue Formation,” *Nature Biotechnology*, Vol. 22, No. 8, 2004, pp. 977-984.
- [71] A. L. Gomes dos Santos, A. Bochot, A. Doyle, *et al.*, “Sustained Release of Nanosized Complexes of Polyethyleneimine and Anti-TGF- β 2 Oligonucleotide Improves the Outcome of Glaucoma Surgery,” *Journal of Controlled Release*, Vol. 112, No. 3, 2006, pp. 369-381.
- [72] C. Gómez-Gaete, N. Tsapis, M. Besnard, A. Bochot and E. Fattal, “Encapsulation of Dexamethasone into Biodegradable Polymeric Nanoparticles,” *International Journal of Pharmaceutics*, Vol. 331, No. 2, 2007, pp. 153-159. <http://dx.doi.org/10.1016/j.ijpharm.2006.11.028>
- [73] T. W. Perkins, B. Faha, M. Ni, *et al.*, “Adenovirus-Mediated Gene Therapy Using Human p21waf-1/cip-1 to prevent Wound Healing in a Rabbit Model of Glaucoma Filtration Surgery,” *JAMA Ophthalmology*, Vol. 120, No. 7, 2002, pp. 941-949. <http://dx.doi.org/10.1001/archophth.120.7.941>
- [74] J. G. Yang, N. X. Sun, L. J. Cui, X. H. Wang and Z. H. Feng, “Adenovirus-Mediated Delivery of p27KIP1 to Prevent Wound Healing after Experimental Glaucoma Filtration Surgery,” *Acta Pharmacologica Sinica*, Vol. 30, No. 4, 2009, pp. 413-423.
- [75] M. G. Spiga and T. Borrás, “Development of a Gene Therapy Virus with a Glucocorticoid-Inducible MMP1 for the Treatment of Steroid Glaucoma,” *Investigative Ophthalmology & Visual Science*, Vol. 51, No. 6, 2010, pp. 3029-3041. <http://dx.doi.org/10.1167/iovs.09-4918>